# TOXICOLOGICAL EFFECTS OF ETHANOLIC EXTRACT OF SEED AND BARK OF PERSEA AMERICANA (LAURACEAE), ON LARVAE AND PUPAE OF AEDES ALBOPICTUS (SKUSE, 1894) (DIPTERA, CULICIDAE)

George Harrison Ferreira de Carvalho<sup>1</sup>

**ABSTRACT:** The toxicological effects of crude ethanolic extracts (CEE) of the seed and bark of *Persea americana* have been analyzed on larvae and pupae of *Aedes albopictus* secondary vector of dengue virus and main vector of chikungunya arboviruses in Asian countries. It was used 20 pupae and 20 larvae for each concentration and for each repetition, both for testing and controls, they were made in water and dimethylsulfoxide (DMSO). Mortality was observed after 24 h exposure of larvae and pupae. The lethal concentrations of the LC<sub>50</sub> and LC<sub>90</sub> (CEE) seed and stem bark of *P. americana* were, respectively, 3.5, 7.4 and 4.2, 8.2 mg/L for larvae, and 75.2, 132.1 and 68.9, 115.5 mg/L for pupae. These results indicate the possibility of using this plant for integrated control of *Ae. albopictus* mosquitoes and other disease vectors. Suggest further investigation of the active compounds through chemical studies.

**KEY WORDS**: Aedes albopictus; Persea americana; pupae; larvae; dengue.

EFEITOS TOXICOLÓGICOS DO EXTRATO ETANÓLICO DE SEMENTE E CASCA DE *PERSEA AMERICANA* (LAURACEAE), EM LARVAS E PUPAS DE *AEDES ALBOPICTUS* (SKUSE, 1894) (DIPTERA, CULICIDAE)

**RESUMO:** Foram analisados os efeitos toxicológicos dos extratos brutos etanólico (e.b.e) da semente e da casca de *Persea americana* sobre larvas e pupas do *Aedes albopictus* vetor secundário do vírus do dengue e principal vetor do arbovirus chikungunya nos países Asiáticos. Utilizaram-se 20 larvas e 20 pupas para cada concentração e para cada repetição, tanto para os testes e quanto para os controles, feitos em água e dimetilsulfóxido (DMSO). A mortalidade foi observada após 24h de exposição das larvas e das pupas. As concentrações letais CL<sub>50</sub> e *CL*<sub>90</sub> dos (e.b.e) de semente e da casca do caule de *P. americana* encontradas foram, respectivamente, de 3,5; 7,4 e 4,2; 8,2 mg/L para larvas, e de 75,2; 132,1 e 68,9; 115,5 mg/L para pupas. Esses resultados indicam a possibilidade de uso integrado dessa planta para controle do *Ae. albopictus* e de outros mosquitos vetores de doenças. Sugerem ainda, a investigação dos compostos ativos através de estudos químicos.

PALAVRAS-CHAVE: Aedes albopictus; Persea americana; pupa; larva; dengue.

<sup>&</sup>lt;sup>1</sup> Enfermeiro. Mestre em Medicina Tropical e Saúde Pública da Universidade Federal de Goiás (IPTSP/UFG). Professor e coordenador do curso de enfermagem da Faculdade União de Goyazes. Email:georgeharrisonfc@gmail.com.

### INTRODUCTION

Dengue is a disease transmitted by mosquitoes of the gender *Aedes sp*, being incriminated several speciess of the subgender Stegomyia (*Aedes aegypti, Aedes albopictus* and *Aedes polynesiensis*), in which the virus was found in natura.<sup>1, 2</sup> Laboratory experiments also showed the susceptibility of other speciess of *Aedes* infected with the virus dengue.<sup>2</sup> The most important vector of dengue, however, is *Ae. aegypti.* The *Aedes albopictus* (Skuse,1894) is a vector secondary importance, <sup>3</sup> on the other hand, in some areas of Indonesia, outbreaks frequently have occurred in the rural parts of the country, where *Ae. albopictus* is the prevalent species.<sup>4</sup> In addition, studies of dengue transmission in villages in Thailand indicated the important function of the *Ae. albopictus* in the transmission of dengue disease.<sup>5, 6</sup> In this region *Ae. albopictus* is also the main vector of the chikungunya virus. A disease that causes symptoms and that has been confused with dengue<sup>7</sup>.

*Ae. albopictus* is of Asian origin, was first described in India, where it spread to almost all countries in that region. It was also introduced in the Americas in 1985 in the southern United States<sup>8,9</sup> and a year later appeared in Brazil in the state of Rio de Janeiro<sup>10,11</sup>. In the same year this species was found in Minas Gerais and São Paulo<sup>12</sup> and in the following year, in Espirito Santo<sup>13</sup>. Some researchers believe that this species entered in Brazil through the sea by trade of iron with Japan<sup>10</sup>. In 2003, only the states of Acre, Amapá, Piauí, Sergipe, Tocantins and Roraima, were non-infested by *Ae. Albopictus*.<sup>14</sup>

In Brazil, this mosquito is found in rural and wild areas, but easily adapts to the urban environment. This vector has anthropophilic preference with moderate attraction zoophilic, its biting is diurnal.<sup>15,16</sup> Its larvae colonize all kinds of natural or artificial containers, showing its ecological valence, adapting easily to the rural and urban environments, and can participate in wild cycles or serve as a link between the sylvatic cycle and man<sup>17</sup>. They are more commonly found in tree holes and overlapping leaf plants such as bromeliads<sup>16,18,19,20</sup>. According to recent studies<sup>21</sup>, this mosquito has a good answer to the life in the cities.

In this context, in urban condition, the *Ae. albopictus* shares the same containers as *Ae. Aegypti*,  $^{22,23}$  presenting a greater population than the latter,  $^{24,25,26}$  which shared in high-density larval, proves to be more resistant than *Ae. aegypti* to over-population  $^{16,27,28,29,30}$ .

In the Americas, there are records of natural infection of *Ae. albopictus* with La Crosse virus, equine encephalitis in Venezuela and in the Unite State of America<sup>31,32,33</sup>. However yet, there has not been any reported case of dengue that has been transmitted by *Ae. albopictus* in any country in the Americas.

Researches made in laboratories in Brazil showed that this species has the power of infection and transmission of yellow fever virus and dengue <sup>34,35,36,37,38,39</sup>.

This fact demonstrates the possibility of dengue outbreaks in places where there is no presence of *Ae. aegypti*. The participation of *Ae. albopictus* in the transmission of yellow fever virus and dengue disease could modify the epidemiology of transmission of these diseases in the Americas<sup>40,41,42,43</sup>.

Therefore, if it's proven that the *Ae. albopictus* is able to transmit dengue and yellow fever, the control or eradication of this mosquito would be very difficult. That is due to its adaptation in various types of environments, including temperate regions at high altitudes. Also, due to the mentioned tolerance, its control becomes more difficult than of *Ae. aegypti* <sup>44,45</sup> - e.g: in a study developed <sup>46</sup> in Campos do Jordão at temperatures between 13°C and 16° C, *Ae. albopictus* infested 10% of the municipality more than the *Ae. aegypti*.

Beside its potential as a vector of dengue and yellow fever, in laboratory experiments the *Ae. albopictus* proved to be efficient in the transmission of *Dirofilaria immitis* in some areas of Japan. This insect has been found naturally infected by *D. immitis*, that has been considered as a vector in that country  $^{47}$ .

Ae. albopictus can be a problem in public health politics. Though, this study sought to present the toxicological effects of ethanolic extracts of seeds and bark of Persea americana and its power insecticide on the Ae. albopictus is providing information that can be useful for planning and executing measures to combat and/or control insect vectors.

Several specialists have emphasized the development of herbal substances for mosquito control <sup>48,49,50,51,52,53,54,55,56,57,58</sup>. Plants with application in the pharmaceutical field have been the most interested research about to develop compounds with insecticidal properties <sup>59</sup>. These compounds originated from plants have received special attention because they are environmental friendly alternatives. Then, other favorable factors have been selective, biodegradable and low environmental impact, giving a great security to the population. Many studies have been done with plants in the search for active molecules against vectors, specially the mosquitoes <sup>60,61,62,63,64,65</sup>.

Recent studies have shown that oil from the seed of *Persea americana*, commonly known as avocado, had insecticide power on 3<sup>rd</sup> instar larvae of the *Ae. aegypti* <sup>66</sup>. The objective of this study was to evaluate the larvicidal and pupicidal effect of the ethanolic extracts of seeds and bark of *P. americana* on *Ae. albopictus*.

### **MATERIALS AND METHODS**

Seeds and bark of *P. americana* were collected in the town of Aparecida de Goiânia, which were brought to the laboratory Bioactivity Plant of the Instituto de Patologia Tropical e Saúde Pública (IPTSP), Universidade Federal de Goiás (UFG) for extraction.

The seeds and bark were placed separately in an oven with forced airflow at 40 °C for drying, then crushed in a knife mill to achieve low grain size and cold percolated. This process consisted of placing about 1000g of the powder obtained from the seeds and bark of the *P. americana* in a beaker with a capacity of 2000 ml,

and added one liter of absolute ethyl alcohol and mixed with a mechanical stirrer until became homogeneous.

Each beaker was covered with aluminum foil to prevent evaporation of the alcohol and a possible interference of the light, remaining for 72 hours. The supernatant was filtered through glass funnel with a disposable paper filter.

The filtrate ones were put in a rotary evaporator and the obtained extracts were placed separately in Petri dishes to be dried up in a laminar flow. After solvent evaporation, the extracts were transferred to an upright freezer at a temperature of -4 ° C to be used later.

The 3<sup>rd</sup> instar larvae and pupae of *Ae. albopictus* used in the tests were created according to the methodology of Silva et al. (1998)<sup>67</sup>, with supply of public water in biological chamber heated to 28 at about 1 °C, humidity of 80 at about 5% and photophase of approximately 12 hours.

The bioessays were performed in another climatic chamber similar of the creation (mentioned above). There were three replicates of each experiment, with their respective controls, the crude ethanol extract (CEE) of seed and bark of P. americana to determine the lethal concentrations,  $LC_{50}$  and  $LC_{90}$  for the larvae and pupae of Ae. albopictus.

The solutions used for testing were prepared weighting the (CEE) seeds and bark of *P. americana* in an analytical balance with accuracy of 0.0001 g. Then they (CEE) were dissolved in DMSO (dimethylsulfoxide) 2% and distilled water, remaining at about an hour until dissolution. Subsequently homogenized in a magnetic stirrer at about 10 minutes, the amount has been adjusted with distilled water. The solutions were prepared 24 hours prior to testing in disposable plastic cups, with 100 ml of each solution.

20 larvae and 20 pupae of 3<sup>rd</sup> instar were used for each bioessay and a similar process to the control group, replacing the solution by distilled water plus DMSO 2%. The larvae and pupae were placed in solutions with the aid of disposable plastic pipettes.

The records of mortality were taken after 24 hours of exposure of larvae and pupae of the extract, were considered dead larvae and pupae totally inert, associated with darkening of the body. The lethal concentrations were made by using the Probit Analysis program developed by (Thomas & Alexandra Sparks 1987).

# **RESULTS AND DISCUSSION**

There has been mortality of larvae and pupae of Ae. albopictus in all repetitions and all concentrations of both extracts. The lethal concentrations  $LC_{50}$  and  $LC_{90}$  found

for 3<sup>rd</sup> instar larvae of *Ae. albopictus* (Table 1) were respectively 3.5 and 7.4 mg/L for the seed extract and 4.2 and 8.2 mg/L for the bark extract of *P. americana*.

The result  $LC_{50}$  and  $LC_{90}$  found to the pupae were 75.2 and 132.1 mg/L for the seed extract and 68.9 and 115.5 mg/L to the bark extract of *P. americana* respectively (Table 1). There was no death neither larvae or pupae in control groups DMSO 2%.

**Table 1.** Susceptibility of larvae third instars and pupae of *Aedes albopictus* to crude ethanol extracts of seeds and bark of the *Persea americana* in the laboratory after 24 hours of exposure.

	SE		BS	
Stage	3 <sup>rd</sup> instar	pupae	3 <sup>rd</sup> instar	pupae
LC <sub>50(mg/L)</sub> CI/95%	3,5 (2,7-4,1)	75,2 (73,0-77,5)	4,2 (3,3-5,4)	68,9 (65,7-70,3)
LC <sub>90(ml/L)</sub> Cl/95%	7,4 (5,5-8,9)	132,1 (129,0- 155,8)	8,2 (6,6-10,3)	115,5 (109,8-120,1)

LC: Lethal concentrations;  $LC_{50}$  lethal concentration necessary to kill 50% of larvae and pupae;  $LC_{90}$  lethal concentration necessary to kill 90% of larvae and pupae; mg/L: milligrams liter; 95%CL - Confidence interval at 95% probability; SE: Seed Extracts; BS: Bark Extracts.

Currently, several studies have been presented with plant extracts for mosquito control, especially with larvicidal action. New research, however, need to be developed to evaluate the effect of these extracts on pupae.

The toxicological action of extracts of seeds and bark of *P. americana* on pupae of *Ae. albopictus* have just been observed at high concentrations compared with the results obtained with the 3<sup>rd</sup> instars larvae, it's probably because the pupae is protected by the last larval molt, hindering the action of the extracts.

This protection was broken possibly with the use of high extract concentrations, as noted in a research by Macchione et al. (2004)<sup>68</sup> with *Codonopsis javanica* on *Ae. albopictus* with 75% mortality of pupae in a concentration of 60.000 mg/L. This lethal concentration was about six hundred times higher than the seeds and bark extracts of *P. americana* at this work.

Nathan et al. (2006)<sup>69</sup> reached respectively 92.3% and 90.9% mortality in 20.000 mg/L. with extracts of leaves and seeds of *Melia azedarach* on *Anopheles stephensi*. Nathan (2007)<sup>70</sup> tested *Eucalyptus tereticornis* on *Anopheles stephensi* and found that 160 mg/L killed 88% of the pupae, this result was higher than those obtained with the (CEE) seeds and bark of *P. americana* at this study.

Researches with other *Culicidae* are presented by Murugan et al. (2007)<sup>71</sup>, who used extracts of *Albizzia amara* and *Ocimum basilicum* that had action on the pupae of *Ae. aegypti* in concentrations of 20.000 to 100.000 mg/L. These concentrations were approximately one thousand times higher than those obtained with the (CEE) seeds and bark of *P. americana*.

The LC<sub>50</sub> of the (CEE) seed *P. americana* to  $3^{rd}$  instar larvae of *Ae. albopictus* in the laboratory was 7.2 mg/L, this concentration was less than that obtained by Leite et al.  $(2009)^{66}$  which result was LC<sub>50</sub> of 8.8 mg / L, using the seed extract of this plant on  $3^{rd}$  instar larvae of *Ae. aegypti*.

Researches about plants larvicidal activity to mosquitos show diversify results with variable lethal concentrations like the essays bellow. In the Republic Philippines, Monzon et al. (1994)<sup>72</sup> obtained results of higher lethal concentrations - through the crude extract dissolved in water of these plants *Lansium domesticum, Azedarach indica, Eucaliptus globosus* and *Codiaeum variegatum* to 3<sup>rd</sup> and 4<sup>th</sup> instars of *Culex quinquefasciatus*. These are CL<sub>90</sub> 37, 28, 35 and 24 g/100 ml of water respectively. This concentration is higher than CL<sub>90</sub> to the 3<sup>rd</sup> instar of *Ae. albopictus* found at this work. Silva *et al.* (1996)<sup>73</sup> and Guimarães *et al.* (2001)<sup>74</sup> have verified the lethal concentrations in (CEE) of stain's bark *Magonia pubescens* of 140 and 150 mg /100 ml of destiled water to *Ae. aegypti* and *Ae. Albopictus* too. Both concentrations are higher than the ones found at this work.

### CONCLUSION

The Crude ethanolic extracts of seeds and bark of *P. americana* at this experiment, showed insecticidal activity against larvae and pupae of *Ae. Albopictus*. These results suggest isolation and purification of active compounds through chemical studies in the hope of obtaining lower lethal concentrations.

## **ACKNOWLEDGEMENTS**

To Instituto de Patologia Tropical e Saúde Pública (IPTSP) and Dr Ionizete Garcia da Silva for allowing the use of Plants Bioactivity's laboratories and Insect's Physiology with his help and wisdom to the development of this study.

## REFERENCES

- 1. Bancroft TL 1906. On the etiology of dengue fever. *Australas Medical Gazette*.25:17-18.
- 2. Rodhain F, Rosen L, Gubler DJ, Kuno G 1997. Mosquito vectors and dengue virus-vector relationships. In: Dengue and dengue hemorragic fever. New York: *CAB International* p. 1-22.
- 3. World Health Organization. Dengue [monography on the Internet] Geneve: WHO; 2002 [acessado 2011 set. 20]. Disponível em: <a href="http://www.who.int/ctd/dengue/burdens.htm">http://www.who.int/ctd/dengue/burdens.htm</a>
- 4. Jumali Sunarto, Gubler DJ, Nalim S, Eram S, Sulianti Saroso J 1979. Epidemic dengue hemorrhagic fever in rural Indonésia. III. Entomological studies. *Am J Trop Med Hyg*;28(4):717-724.

- 5. Eamchan P, Nisalak A, Foy HM, Chareonsook OA. Epidemiology and control of dengue virus infections in Thai villages in 1987. *Am J Trop Med Hyg* 1989;41(1):95-101.
- 6. Chareonsook DC, Yap HH 1977. Chemical methods for the control of vector and pests of public health importance. Geneve: WHO/CTD/ WHOPES/97.2.
- 7.Townson H, Nathan MB 2008. Resurgence of Chikungunya. *Trans R Soc Trop Med Hyg 102*: 308-309.
- 8. Sprenger D, Wuithranyagool T 1986. The discovery e distribution of *Aedes albopictus* in Harris Country, Texas. *J Am Mosq Control Assoc 2*: 217-219.
- 9. Rai KS 1991. Aedes albopictus in the Americas. Annu Rev Entomol 36: 459-484.
- 10. Forattini OP 1986. Identificação de *Aedes (Stegomyia) albopictus* (Skuse) no Brasil. *Rev Saúde Pública 20*: 244-245.
- 11. Funasa Fevereiro de 2001 Histórico da presença do *Aedes aegypti* e *Aedes albopictus* no Brasil, p.23-24.
- 12. Brito M, Marques GRAM, Marques CCA, Tubaki RM 1986. Primeiro encontro de *Aedes (Stegomyia) albopictus* (Skuse) no Estado de São Paulo. *Rev Saúde Pública* 20(6): 489.
- 13. Ferreira Neto JA, Lima MM, Aragão MB 1987. Primeiras observações sobre o *Aedes albopictus* no Estado do Espiríto Santo, Brasil. *Cad Saúde Pública 3*(1): 56-61.
- 14. Santos RLC 2003. Atualização da distribuição de *Aedes albopictus* no Brasil (1997-2002). *Rev Saúde Pública 37*(5): 671-673.
- 15. Marques GRAM, Gomes AC 1997. Comportamento antropofílico de *Aedes albopictus* (Skuse) (Diptera: Culicidae) na região do Vale do Paraíba, Sudeste do Brasil. *Rev Saúde Pública 31*(2): 125-130.
- 16. Gomes AC, Forattini OP, Kakitani I, Marques GRAM, Marques CCA, Marucci D, Brito M 1992. Microhabitats de *Aedes albopictus* (Skuse) na região do Vale do Paraíba, Estado de São Paulo. *Rev Saúde Pública 26*(2): 108-18.
- 17. PAHO Pan American Health Organization 1995. Biology, disease relationships, and control of *Aedes albopictus*. Washington (DC). *Technical Paper*, 42.
- 18. Natal D, Urbinatti PR, Taipo-Lagos, Cereti-Júnior W, Diederichsen ATB, Souza RG, Souza RP 1997. Encontro de *Aedes (Stegomyia) albopictus (Skuse)* em Bromeliaceae na periferia de São Paulo, SP, Brasil. *Rev Saúde Pública 31*(5): 517-518.

- 19. Forattini OP, Marques GRAM, Kakitani I, Brito M, Sallum MAM 1998. Significado epidemiológico dos criadouros de *Aedes albopictus* em bromélias. *Rev Saúde Pública 32*(2): 186-188.
- 20. Marques GRAM, Santos RC, Forattini OP 2001. *Aedes albopictus* em bromélias de ambiente antrópico no Estado de São Paulo, Brasil. *Rev Saúde Pública 35*(3): 243-248.
- 21. Forattini OP, Kakitani I, Santos RC, Kobayashi KM, Ueno HM, Fernandez Z 2000. Comportamento de *Aedes albopictus* e de *Ae. scapularis* adultos (Diptera: Culicidae) no Sudeste do Brasil. *Rev Saúde Pública 34*(5): 461-467.
- 22. Brito M, Forattini OP 2004. Produtividade de criadouros de *Aedes albopictus* no Vale do Paraíba, SP, Brasil. *Rev Saúde Pública 38*(2): 209-215.
- 23. Silva VC, Scherer PO, Falcão SS, Alencar J, Cunha SP, Rodrigues IM, Pinheiro NL, 2006. Diversidade de criadouros e tipos de imóveis freqüentados por *Aedes albopictus* e *Aedes aegypti. Rev Saúde Pública 40*(6): 1106-1111.
- 24. Honório NA, Lourenço-de-Oliveira R 2001. Freqüência de larvas e pupas de *Aedes aegypti* e *Aedes albopictus* em armadilhas, Brasil. *Rev Saúde Pública 35*(4): 385-391.
- 25. Almeida OS, Ferreira AD, Pereira VL, Fernandes MG, Fernandes WD 2006. Distribuição espacial de *Aedes albopictus* na região sul do Estado de Mato Grosso do Sul. *Rev Saúde Pública 40*(6): 1094-1100.
- 26. Honório NA, Cabello PH, Codeço CT, Lourenço-de-Oliveira R 2006. Preliminary data on the performance of *Aedes aegypti* and *Aedes albopictus* immatures developing in water-filled tires in Rio de Janeiro. *Mem Inst Oswaldo Cruz 101*(2): 225-228.
- 27. Gomes AC, Gotlied SLD, Marques CCA, Paula MB, Marques GRAM 1995. Duration of larval and pupal development stages of *Aedes albopictus* in natural and artificial containers. *Rev Saúde Pública 29*(1): 09-15.
- 28. O'Meara GF, Evans LF, Gettman ADJ, Cuda JP 1995. Spread of *Aedes albopictus* and decline of *Ae. aegypti* (Diptera: Culicidae) in Florida. *J Med Entomol* 32(5): 554- 562.
- 29. Silva MAN, Calado DC, Tissot AC, Chrestani M 2001. Biologia de imaturos e adultos de *Aedes albopictus* sob condições de laboratório e ecologia de Culicidae em área de mata de Curitiba, PR. *Inf Epidemiol* 10(1): 17-19.
- 30. Barreira R 1996. Competition and resistance to starvation in larvae of container-in habiting *Aedes* mosquitoes. *Ecol Entomol* 21: 117-127.
- 31. Mitchell C, Niebylski M, Smith G, Karabatsos N, Martin D, Mutebi JP, Craig GB, Mahler M 1992. Isolation of Eastern equine encephalitis from *Ae. albopictus* in Florida. *Science 257*: 526-527.

- 32. Moore CG, Mitchell CJ 1997. *Aedes albopictus* in the United State: ten-year presence and public health implications. *Emerg Infec Dis 3*: 329-334.
- 33. Gerhardt RR, Gottfried KL, Apperson CS, Davis BS, Erwin PC, Smith AB, Panella NA, Powell EE, Nasci RS 2001. First isolation of La Crosse Virus from naturally infected *Aedes albopictus*. *Emerg Infec Dis* 7(5): 807-811.
- 34. Savage HM, Ezike VI, Nwankwo ACN, Spiegel R, Miller BR 1992. First record of breeding populations of *Aedes albopictus* in Continental Africa: implications for arboviral transmission. *J Am Mosq Control Assoc 8*: 101-103.
- 35. Serufo JC, Montes de Oca H, Tavares V, Souza AM, Rosa RV, Jamal MC 1993. Isolation of dengue virus type 1 from larvae of *Aedes albopictus* in Campos Altos City, State of Minas Gerais, Brazil. *Mem Inst Oswaldo Cruz 88*(3): 503-504.
- 36. Gomes AC, Bitencourt MD, Natal D, Pinto LSP, Mucci LF, De Paula MB, Urbinatti PR, Barata JMS 1999. *Aedes albopictus* em área rural do Brasil e implicações na transmissão de febre amarela silvestre. *Rev Saúde Pública 33*(1): 95-97.
- 37. Lourenço-de-Oliveira R, Vazeille M, Filippis AMB, Failloux AB 2003. Large gentic differentiation and low variation in vector competence for dengue and yellow fever viruses of *Aedes albopictus* from Brazil, the United States, and the Cayman Islands. *Am J Trop Med Hyg 69*: 105 -114.
- 38. Guedes DRD 2006. *Epidemiologia molecular do Aedes albopictus (Diptera: Culicidae)*. Dissertação de Mestrado. Fundação Oswaldo Cruz, Recife.
- 39. Martins VEP, Martins MG, Araújo JMP, Silva LOR, Monteiro HAO, Castro FC, Vasconcelos PFC, Guedes MIF 2006. Primeiro registro de *Aedes (Stegomyia) albopictus* no Estado do Ceará, Brasil. *Rev Saúde Pública 40*(4): 737-739.
- 40. Miller BR & Ballinger ME 1988. *Aedes albopictus* mosquitoes introduced into Brazil: vector competence for yellow fever and dengue viruses. *Trans R Soc Trop Med Hyg 82*: 476-477.
- 41. Mitchell CJ & Miller BR 1990. Vertical transmission of Dengue Viruses vy strains of *Aedes albopictus* recently introduced into Brazil. *J Am Mosq Control Assoc 6*(2): 251-253.
- 42. Chiaravalloti Neto F, Dibo MR, Barbosa AAC, Battigaglia M 2002. *Aedes albopictus* (S) na região de São José do Rio Preto, SP: estudo da sua infestação em área já 20 ocupada pelo *Aedes aegypti* e discussão de seu papel como possível vetor de dengue e febre amarela. *Rev Soc Bras Med Trop 35*(4): 351-357.
- 43. Castro MG, Nogueira RMR, Schatzmayr HG, Miagostovich MP, Lourenço-de-Oliveira R 2004. Dengue vírus detection by using reverse transcription polymerase chain reaction in saliva and progeny of experimentally infected *Aedes albopictus* from Brazil. *Mem Inst Oswaldo Cruz 99*(8): 809-814.

- 44. Hawley WA 1988. The biology of *Aedes albopictus*. *J Am Mosq Control Assoc 4*: 2-39.
- 45. Marques GRAM, Gomes AC 1997. Comportamento antropofílico de *Aedes albopictus* (Skuse) (Diptera: Culicidae) na região do Vale do Paraíba, Sudeste do Brasil. *Rev Saúde Pública 31*(2): 125-130.
- 46. Glasser CM, Gomes AC 2002. Clima e sobreposição da distribuição de *Aedes aegypti* e *Aedes albopictus* na infestação do Estado de São Paulo. *Rev Saúde Pública 36*(2): 166-172.
- 47. Konishi E 1989. *Culex tritaeniorhynchus* and *Aedes albopictus* (Diptera: Culicidae) as natural vectors of *Dirofilaria immitis* (Spirurida: Filariidae) in Miki City, Japan. *J Med Entomol* 26(4): 294-300.
- 48. Amer A, Mehlhorn H 2006b. Persistency of larvicidal effects of plant oil extracts under different storage conditions. *Parasitol Res* 99: 473-477.
- 49. Champakaew D, Choochote W, Pongpaibul Y, Chaithong U, Jitpakdi A, Tuetun B, Pitasawat B 2007. Larvicidal efficacy and biological stability of a botanical natural product, zedoary oil-impregnated sand granules, against *Aedes aegypti* (Diptera, Culicidae). *Parasitol Res* 100: 729-737.
- 50. Cheng SS, Huang CG, Chen YJ, Yu JJ, Chen WJ, Chang ST 2009. Chemical compositions and larvicidal activities of leaf essential oils from two eucalyptus speciess. *Bioresource Technol* 1: 452-456.
- 51. Mullai K, Jebanesan A, Pushpanathan T 2008. Effect of bioactive fractions of *Citrullus vulgaris* Schrad. leaf extract against *Anopheles stephensi* and *Aedes aegypti. Parasitol Res* 102: 951-955.
- 52. Pavela R 2008. Larvicidal effects of various Euro-Asiatic plants against *Culex quinquefasciatus* Say larvae (Diptera: Culicidae). *Parasitol Res* 102: 555-559.
- 53. Pushpanathan T, Jebanesan A, Govindarajan M 2008. The essential oil of *Zingiber officinalis* Linn (Zingiberaceae) as a mosquito larvicidal and repellent agent against the filarial vector *Culex quinquefasciatus* Say (Diptera: Culicidae). *Parasitol Res* 102: 1289-1291.

- 54. Rahuman AA, Gopalakrishnan G, Venkatesan P, Geetha K 2008. Larvicidal activity of some Euphorbiaceae plant extracts against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitol Res* 102: 867-873.
- 55. Schmutterer H 1990. Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. *Annu Rev Entomol* 35: 271-297.
- 56. Silva OS, Prophiro JS, Rossi JCN, Kanis LA, Romão PRT, Blazius RD 2006. Larvicidal effect of andiroba oil *Carapa guianensis* (Meliaceae) against *Aedes aegypti* (Diptera: Culicidae). *J Am Mosq Control Assoc* 22: 699-701.
- 57. Silva JS, Mariano FZ, Scopel I 2008. Ano dengue no Brasil e as políticas de combate ao *Aedes aegypti*: Da tentativa de erradicação as políticas de controle. *Hygeia* 3: 163-175.
- 58. Simas NK, Lima EC, Conceição SR, Kuster RM, Filho AMO 2004. Produtos naturais para o controle da transmissão da dengue atividade larvicida de *Myrozylon balsamum* (óleo vermelho) e de terpenóides e fenilpropanóides. *Quim Nova* 27: 46-49.
- 59. Souza SS, Silva IG, Silva HHG 2010. Associação entre incidência de dengue, pluviosidade e densidade larvária de *Aedes aegypti*, no Estado de Goiás. *Rev Soc Bras Med Trop* 43: 152-155.
- 60. Aguilera L, Navarro A, Tacoronte JE, Leyva M, Marquetti MC 2003. Efecto letal de myrtaceas cubanas sobre *Aedes aegypti* (Diptera: Culicidae). *Rev Cubana Med Trop 55*: 100-104.
- 61. Cavalcanti ESB, Morais SM, Lima MAA, Santana EWP 2004. Larvicidal Activity of Essential Oils from Brazilian Plants against *Aedes aegypti* L. *Mem Inst Oswaldo Cruz* 99: 541-544.
- 62. Magadula JJ, Innocent E, Otieno JN 2009. Mosquito larvicidal and cytotoxic activities of 3 *Annona* speciess and isolation of active principles. *J Med Plants Res* 3(9): 674-680.

- 63. Silva H H G, Geris R, Silva IG, Rodrigues FE 2007. Larvicidal activity of oil-resin fractions from the brazilian medicinal plant *Copaifera reticulata. Rev Soc Bras Med Trop* 40:264-267.
- 64. Silva IG, Guimarães VP, Lima CG, Silva HHG, Elias CN, Mady CM, Silva VVM, Nery AP, Rocha KR, Rocha C, Isac E 2003. Efeito larvicida e toxicológico do extrato bruto etanólico da casca do caule de *Magonia pubescens* sobre *Aedes aegypti* (Diptera, Culicidae), em criadouros artificiais. *Rev Patol Trop 32*: 73-86.
- 65. Simas NK, Lima EC, Conceição SR, Kuster RM, Filho AMO 2004. Produtos naturais para o controle da transmissão da dengue atividade larvicida de *Myrozylon balsamum* (óleo vermelho) e de terpenóides e fenilpropanóides. *Quim Nova 27*: 46-49.
- 66. Leite JJ, Brito EH, Cordeiro RA, Brilhante RS, Sidrim JJ, Bertini LM, Morais SM, Rocha MF 2009. Chemical composition, toxicity and larvicidal and antifungal activities of *Persea americana* (avocado) seed extracts. *Rev Soc Bras Med Trop* 42: 110-113.
- 67. Silva HHG, Silva IG, Lira KS 1998. Metodologia de criação, manutenção de adultos e estocagem de ovos de *Aedes aegypti* (Linnaeus, 1762) em laboratório. *Rev Patol Trop 27*: 53-63.
- 68. Macchioni F, Carugini S, Cecchi F, Siciliano T, Braca A, Cioni P, Morelli I 2004. Aqueous extract of *Codonopsis javanica* against larval and pupal stages of *Aedes albopictus*. Annali *della Facoltà di Medicina Veterinária*. 57:215-20.
- 69. Nathan SS, Savitha G, George DK, Narmadha A, Suganya L, Chung PG 2006. Efficacy of *Melia azedarach* L. extract on the malarial vector *Anopheles stephensi* Liston (Diptera: Culicidae). Bioresour Technol. 97:1316–23.
- 70. Nathan SS 2007. The use of *Eucalyptus tereticornis* Sm. (Myrtaceae) oil (leaf extract) as a natural larvicidal agent against the malaria vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Bioresour Technol.* 98:1856–60.
- 71. Murugan K, Murugan P, Noortheen A 2007. Larvicidal and repellent potential of *Albizzia amara* Boivin and *Ocimum basilicum* Linn against dengue vector, *Aedes aegypti* (Insecta:Diptera:Culicidae). *Bioresour Technol.* 98:198–201.
- 72. Monzon RB, Lucson LLC, Morales AS, Mutuc FES 1994. Larvicidal potencial of five phiippine plants against *Aedes aegypti* (Linnaeus) and *Culex quinquefasciatus* (Say). *Southteast Asian J Trop Med Public Health 25*: 755-759.

- 73. Silva IG, Santos AH, Ferri PH, Alves FBN, Melo RQ, Peixoto L, Silva HHG, Elias CN, Isac E, Lira KS, Camargo MF 1996. Ação larvicida de extrato bruto etanólico de *Magonia pubescens St. Hil* (tingui-do-Cerrado), sobre o *Aedes aegypti (Lin.)* em laboratório. *Rev Pat Trop.* 25: 51-59.
- 74. Guimarães VP, Silva IG, Silva HHG,Rocha C 2001. Atividade larvicida do extrato bruto etanólico da casca do caule de *Magonia pubescens* St. Hil. sobre o *Aedes albopictus* (Skuse, 1894) (Diptera, Culicidae). *Rev Patol Trop. 30*: 243-249.